



**ANNA KOZIOROWSKA<sup>1</sup>, KATARZYNA STATS<sup>2</sup>,  
MARIA ROMEROWICZ-MISIELAK<sup>3</sup>**

## **The influence of environmental factors on metabolic activity of cancer cells**

<sup>1</sup> Doktor inżynier, Uniwersytet Rzeszowski, Wydział Matematyczno-Przyrodniczy, Katedra Inżynierii Komputerowej, Polska

<sup>2</sup> Inżynier, studentka, Uniwersytet Rzeszowski, Instytut Biotechnologii Stosowanej i Nauk Podstawowych, Katedra Fizjologii i Rozrodu Zwierząt, Polska

<sup>3</sup> Doktor, Uniwersytet Rzeszowski, Instytut Biotechnologii Stosowanej i Nauk Podstawowych, Katedra Fizjologii i Rozrodu Zwierząt, Polska

### **Abstract**

The paper presents the results of viability of breast cancer cells under the influence of electromagnetic field. MCF-7 cell line was subjected to an electromagnetic field with a frequency of 5 Hz, 60 Hz and 120 Hz and an MTT assay was performed immediately after the influence of the field and after 24 hours. No statistical difference was demonstrated in cell viability immediately after exposure to EMF, and there are demonstrated differences in the case of field frequencies of 5 Hz and 120 Hz within 24 hours after exposure.

**Key words:** cells viability, Electromagnetic field.

---

### **Introduction**

Electromagnetic field (EMF) accompanies people in all areas of life. It is one of environmental factor which influence our everyday life. It is the space in which electric and magnetic energy permeate each other. These interrelationships are described by Maxwell's equations.

The electric field is directed perpendicular to the Earth. The value of the electric field strength depends on the weather conditions. During the good weather it is approx. 130 V/m, during dense fog to 2 kV/m, and during a storm it can be up to 20 kV/m. The magnetic field does not depend on atmospheric phenomena and its intensity reaches 40 A/m. [Siemiński 1994; Kudowski et al. 1997].

Sun and Earth produce the natural electromagnetic field and artificial fields are produced by a growing number of everyday devices. The sources of natural magnetic field are:

1. Geomagnetic field of the Earth (currently the magnetic south pole is located approx. 7.3° from the north pole of the Earth and the magnetic south pole of approx. 27° from the north pole of the Earth).

2. Atmospheric Phenomena (eg. Electrical discharges).
3. The fields coming from space (the most powerful source of the magnetic field is Jupiter and the solar radiation).
4. Ocean and sea tides.

The most widespread artificial sources of EMF are transmission lines and all household appliances supplied from industrial network. These devices are the source of the fields of extremely low frequency range of 50–60 Hz. Among the household appliances are also those which emit fields of higher frequencies, such as microwave ovens or mobile phones often worn in close proximity to the human body. These devices operate at high frequencies above 300 MHz.

Both – the electric the magnetic fields and are used in medicine. There are applied field of the entire frequency spectrum – 0–300 Hz. They are used in the diagnosis (eg. Magnetic resonance imaging), physiotherapy (terapuls, diathermy, treatment of fixed and modulated magnetic field) or interventional medicine (eg. Electrosurgical units).

EMF affecting the environment and human as any physical factor influence on living matter. The impact depends on the method of the field generating and from the time of exposure and frequency range. The electromagnetic field shows broad spectrum of action on living tissues. The mechanisms of these interactions are studied for many years, but are not fully understood.

Depending on the frequency, activity extends from stimulation of excitable tissues such as nerves, muscles and heart [Polk 1995; Palti, 1966] by the stimulation of bone growth and accelerate of fracture healing [Besset, 1985] to use it for the thermal ablation of tumors using electromagnetic waves of a radio frequency. The electric field of an intermediate-frequency (> 10 kHz to MHz) was often considered as not exerting a biological effect [Elson 1995], and hence, medical application, a few non-thermal cell effects which has been observed [Zimmerman et al. 1981; Holzapfel et al. 1982; Pawlowski et al. 1993].

Until 1970 it was thought that exposure to electromagnetic fields is completely safe to people and does not lead to the formation of any damage or disrupt the functioning of tissues. Just in 1979, control clinical studies demonstrated the relationship between the place of residence in the vicinity of power networks with the number of cases of cancer [Wertheimer, Lepper 1979]. Since then, subsequent studies confirmed the relationship between raised risk of developing certain types of cancer, especially in children, and the constant staying in the vicinity power networks [London et al. 1991; Savitz et al. 1988; Feychting, Ahlbom 1993; Tomenius 1985].

One of the cases of examined cancer was breast cancer. The risk of breast cancer is significantly higher in the industrial urban areas, such as northern Europe and North America than in less developed areas, such as Africa and Asia [Stevens 1987; Stevens, Davis 1996]. In 1978, Cohen and his colleagues sug-

gested that the reduction of production of melatonin by the pineal gland can raise estrogen levels in the circulation, stimulate the proliferation of breast tissue and can lead to breast cancer [Cohen et al. 1978]. Cohen and colleagues hypothesized that the environmental light can be a factor which may lead to lower production of melatonin.

The last decade has brought a number of *in vitro* and *in vivo* studies, that have documented the antitumor effect of an alternating electric field [Kirson et al. 2007; Zimmerman et al. 2012], including the intermediate-frequency low-intensity (100–300 kHz) alternating electric field and the magnetic field amplitude-modulated slightly lower frequencies (0.1 Hz to 114 kHz) [Barbault et al. 2009].

Zimmermann et al. [2013] showed that the antitumor effect was achieved in a specific (for the type of tumor), frequency modulation, and demonstrated inhibition of proliferation and disruption of the mitotic spindle when exposed to an alternating electric field [Kirson et al. 2007; Zimmerman et al. 2012]. Moreover, the bridging important aspects of apoptosis [Fang et al. 1998; Silva et al. 1996] with an extremely low frequency (ELF) pulsed-gradient magnetic field, Zhang [Zhang et al. 2002] shows that can not only induce, but also can block the development of neovascularization required for the nutrition (blood supply to the tumor).

The hypothesis of a compound of the electromagnetic field with breast cancer is based on experimental evidence, that the light and electromagnetic fields of extremely low frequency affects the production of melatonin by the pineal gland, thus affecting the mammalian carcinogenesis in laboratory studies [Stevens 1987; Stevens, Davis 1996; Preston-Martin 1996]. Biological plausibility of the relationship between EMF and breast cancer is associated (conjugated) with unexplained high rate of breast cancer cases in some industrialized urban areas, suggesting that further investigations are warranted.

## **Materials and Methods**

### **Cell culture**

The research model used in this experiment was stabilized, derived from a human breast cancer cell line MCF-7. The cells were cultured in DMEM medium supplemented with 10% fetal calf serum (FBS). The culture was carried out at 37°C under 95% humidity and 5% CO<sub>2</sub> concentration in the air. All experiments were performed between 3 and 7 passage.

### **Exposure of MCF-7 cells to electromagnetic fields**

The dynamic magnetic field generator (Magneris, Astar) was used in this study. The distributions of EMF was determined by Astar using: magnetic field meter GM04 (Hirst Magnetic Instruments, UK), Hall effect sensor type A1321

(Allegro MicroSystems), TDS1002B oscilloscope (Tektronix), BM515X digital multimeter (BRYMEN). Apparatus can generate low-frequency electromagnetic field in the range from 2 to 120 Hz with sinusoidal, triangular and rectangular shape. Magnetic field distribution inside the two-part flat applicator gives the opportunity to carry out tests for different values of magnetic induction (in the study from 2 to 6 mT). The shape of the magnetic field was directly dependent on the shape of the current passing through the solenoid (in the study sinusoidal shape, frequency 5 Hz, 60 Hz and 120 Hz).

Experiments were carried out on 96 well culture plates. 24 hours before the experiment, the MCF-7 cells were seeded on a plate at a density of 1000 cells/well. The cells were exposed to EMF by 2 hours. There were taken cells untreated by EMF as a control.

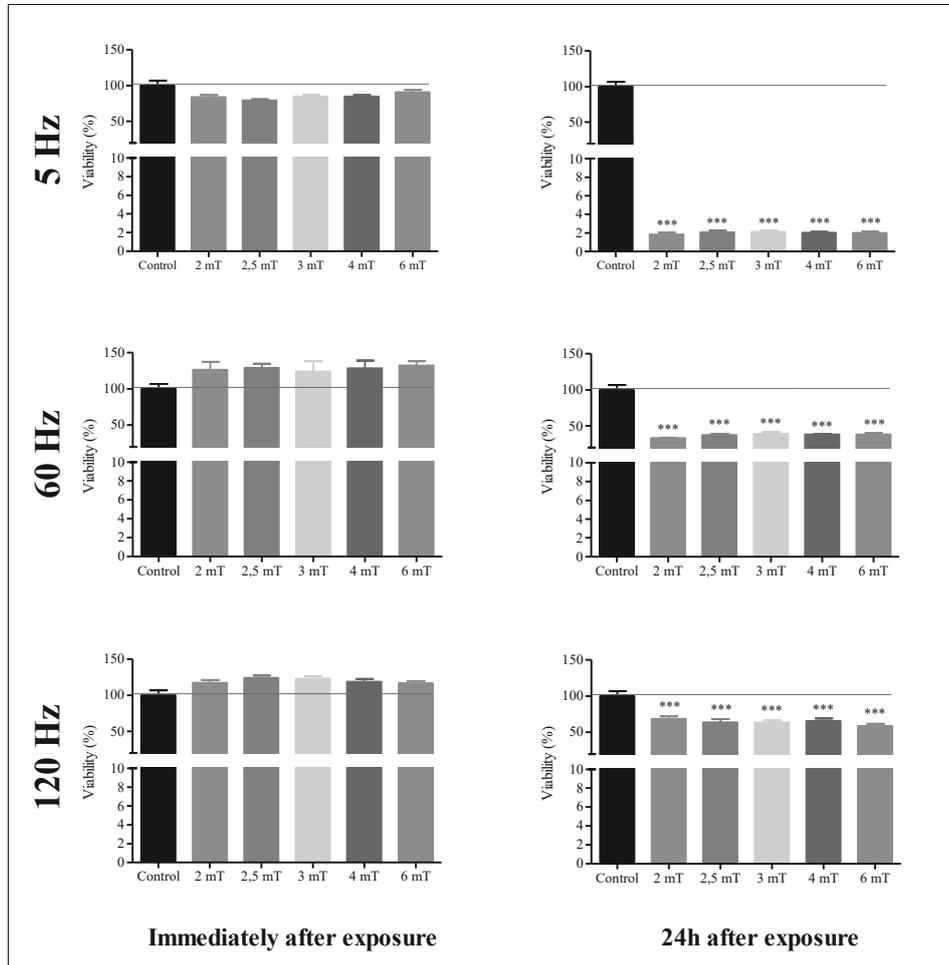
#### **Estimation of cell viability – MTT assay**

Metabolic activity was determined by quantitative colorimetric MTT assay (Sigma-Aldrich). The principle of the test is the reduction of yellow dissolved 3-(4,5-dimethylthiazol-2-yl) -2,5-diphenyltetrazolium bromide to formazan blue, due to the action of succinate dehydrogenase. It is an enzyme functioning only in cells with intact mitochondrial oxidative activity, and therefore the MTT assay is used for the analysis of live cells [Dzitko et al. 2010].

The cells viability was estimated immediately after the two-hour exposure to an electromagnetic field after 24 hours. There was added a solution of 50 µl of MTT reagent at concentration of 5 mg/ml to each well (Sigma-Aldrich; Niemcy). The cultures were incubated for 4 hours at 37°C, 95% humidity and at concentration of 5% CO<sub>2</sub> in the air. After incubation, the precipitated crystals of formazan were dissolved. The liquid from the crystal was removed by pipetting, and then 100 µl of solvent formazan (4 mM HCl, 0.1% Nanodet P-40 in isopropanol) was added to each well and gently stirred for 15 minutes to dissolve the crystals. The absorbance of the solution was measured spectrophotometrically at 492 nm using a Victor plate reader (Perkin Elmer). The results are shown as the percentage of viable cells in relation to the control.

#### **Results**

Immediately after completion of the exposure to electromagnetic fields with frequency of 5 Hz, 60 Hz and 120 Hz there is not shown significant changes in MCF-7 cell viability. 24 hours after exposure to an electromagnetic field with a frequency of 5 Hz and at a magnetic induction of 2 to 6 mT, cells viability significantly decreased ( $P \leq 0.001$ ) in relation to the control. After 24 hours from the completion of the exposure to an electromagnetic field with a frequency of 120 Hz, there was a significant ( $P \leq 0.001$ ) increase in MCF-7 cell viability compared to the control. The results of studies are shown at Fig. 1.



**Fig. 1.** The viability of MCF-7 breast cancer cells under the influence of electromagnetic field with frequency of 5 Hz, 60 Hz and 120 Hz immediately after exposure on EMF and 24 hours after exposure. Data are shown as the mean  $\pm$  SEM, \*\*\* $P \leq 0.001$

## Discussion

In recent years greatly increased interest in the potential use of the electromagnetic field in anticancer therapy [Barbault et al. 2009; Blackman 2012; Cameron et al. 2005; Elson 2009; Zimmerman et al. 2012]. Numerous scientific reports concerning the particular sensitivity of tumor cells to electromagnetic fields of extremely low frequency contributed to this [Zimmerman et al. 2012, Crocetti et al. 2011; Ruiz-Gómez, Martinez-Morillo 2005; Yamaguchi et al. 2006]. At the same time exposure to ELF-EMF was found to be neutral or even beneficial to normal cells [Elson 2009; Repacholi, Greenebaum 1999]. The aim

of this study was to evaluate the effect of electromagnetic fields of extremely low frequency on the viability of MCF-7 breast cancer cells. The analysis was carried out *in vitro*. After 24-hour of incubation, cells were exposed to an electromagnetic field with a frequency of 5 Hz, 60 Hz and 120 Hz and a magnetic induction of 2–6 mT. Cells viability was estimated immediately after completion of exposure to EMF and after 24 hours.

There is demonstrated the significant effect of the electromagnetic field in the studied range of frequencies on the viability of MCF-7 cells. After 24 hours of incubation after the end of 2-hour exposure to EMF, breast cancer cells viability, measured by the MTT assay, was significantly lower in relation to the control (Fig. 1). The effectiveness of electromagnetic fields was dependent on their frequencies, there were no significant differences related to the value of a magnetic induction (2–6 mT). The highest decrease in viability of MCF-7 cells were observed for cell exposure to EMF with frequency of 5 Hz. The similar test results obtained Crocetti et al. [2013], showing the sensitivity of MCF-7 breast cancer cells, measured by the rate of proliferation to an electromagnetic field in the parameters of EMF and exposure time dependent manner. At the same time they did not observe the effect of the cytotoxic effects of EMF on the healthy breast cells (MCF-10). In the scientific papers there are number of studies proving the tumor cell proliferation inhibitory action of the electromagnetic field with the frequency range from 20 Hz to 100 Hz [Buckner et al. 2015; Zhang et al. 2012; Yan et al. 2010; Crocetti et al. 2013]. Studies on the biological effects of electromagnetic fields action relate mainly to its frequency of 50 Hz and 60 Hz, because they are the most common and correlate with environmental exposure (eg. power lines) [Tomitsch, Dechant 2015]. The molecular mechanism of action of the electromagnetic field to the cells and the selectivity induced effects is not fully understood, and required further studies. There is a few pathways on which the EMF may affect the viability and proliferation of cells. One of them is the induction of apoptosis, the programmed cell death that is dependent on oxygen free radicals, which level rises as a result of an electromagnetic field influence. Some studies have shown increased expression of HSP70, a marker of cellular stress response, as a result of stimulation by EMF [Lin et al. 2001; Takalov, Gutzeit 2004]. Also it is speculated that the efficiency of the electromagnetic field antitumor therapy is a result of change (increase) of the level of intracellular calcium ( $\text{Ca}^{2+}$ ) or specific signaling pathways [Wolf et al. 2005; Vijayalaxmi, Prihoda 2009; Simko 2007; Sadeghipour et al. 2012]. Exposure to EMF causes changes in cell membrane integrity allow the influx of  $\text{Ca}^{2+}$  into the cell through voltage-gated calcium channels T [Stratton et al. 2013; Saliev et al. 2014]. However, many tumor cell lines, among which there are the cell lines derived from breast cancer, MCF-7 and MDA-MB-231, are characterized by a unique overexpression of this type of ion channel which was not seen in nor-

mal cells [Taylor et al. 2008a; Capiod 2011; Taylor et al. 2008b; Ohkubo, Yamazaki 2012].

In summary, the observations made in this study are consistent with the idea of having the specific properties of the electromagnetic field, which can alter the functioning of biological systems. Thus, a selective mechanism for killing tumor cells opens the possibility of using a technology based on the electromagnetic field to the preferential destruction of breast cancer cells under clinical conditions.

### Acknowledgements

The study was carried out with the use of equipment granted by the project “Centre of Applied Biotechnology and Basic Sciences” supported by the Operational Programme “Development of Eastern Poland 2007-2013”. No. POPW.01.03.00-18-018/09.

Part of the research was carried out with funding from the municipal office Kolbuszowa.

### Literature

- Barbault A., Costa F.P., Bottger B., Munden R.F., Bomholt F. et al. (2009). *Amplitude-Modulated Electromagnetic Fields for the Treatment of Cancer: Discovery of Tumor-Specific Frequencies and Assessment of a Novel Therapeutic Approach*, „J Exper Clin Cancer Res” no. 28.
- Besset C.A. (1985), *The Development and Application of Pulsed Electromagnetic Fields (PEMFs) for Ununited Fractures and Arthrodeses*, „Clin Plast Surg” no. 12.
- Blackman C.F. (2012), *Treating Cancer with Amplitude-Modulated Electromagnetic Fields: A Potential Paradigm Shift, Again?*, „Br J Cancer” no. 106.
- Buckner C.A., Buckner A.L., Koren S.A., Persinger M.A., Lafrenie R.M. (2015), *Inhibition of Cancer Cell Growth by Exposure to a Specific Time-Varying Electromagnetic Field Involves T-type Calcium Channels*, „PLoS One” no. 10(4).
- Cameron I.L., Sun L.Z., Short N., Hardman W.E., Williams C.D. (2005), *Therapeutic Electromagnetic Field (TEMF) and Gamma Irradiation on Human Breast Cancer Xenograft Growth, Angiogenesis and Metastasis*, „Cancer Cell Int” no. 5.
- Capiod T. (2011), *Cell Proliferation, Calcium Influx, and Calcium Channels*, „Biochimie” no. 93.
- Cohen M., Lippmann M., Chabner B. (1978), *Role of Pineal Gland in Aetiology and Treatment of Breast Cancer*, „Lancet” no. 2.
- Crocetti S., Piantelli F., Leonzio C. (2011), *Selective Destabilization of Tumor Cells with Pulsed Electric and Magnetic Sequences: A Preliminary Report*, „Electromagn Biol Med” no. 30.
- Crochetti S., Beyer C., Schade G., Egli M., Frohlich J., Franco-Obregon A. (2013), *Low Intensity and Frequency Pulsed Electromagnetic Fields Selectively Impair Breast Cancer Cell Viability*, „PLOS ONE” no. 8.
- Dzitko K., Dudzińska D., Grzybowski M., Długońska H. (2010), *The Utility of MTT and XTT Colorimetric Tests in the Studies Conducted in vitro with Toxoplasma gondii tachyzoites*, „Wiad Parazytol.” no. 56(2).

- Elson E. (1995), *Biologic Effects of Radiofrequency and Microwave Fields in vivo and in vitro Experimental Results* [w:] J.D. Bronzino (red.), *The Biomedical Engineering Handbook*, Boca Raton, FL.
- Elson E.I. (2009), *The Little Explored Efficacy of Magnetic Fields in Cancer Treatment and Postulation of the Mechanism of Action*, „Electromagn Biol Med” no. 28.
- Fang M., Zhang H.Q., Xue S.B. (1998), *Role of Calcium in Apoptosis of HL-60 Cells Induced by Harring Tonine*, „Science in China” Ser C 41.
- Feychting M., Ahlbom A. (1993), *Magnetic Fields and Cancer in Children Residing Near Swedish High-Voltage Power Lines*, „Am J Epidemiol.” no. 138.
- Holzappel C., Vienken J., Zimmermann U. (1982), *Rotation of Cells in an Alternating Electric Field: Theory and Experimental Proof*, „J Membr Biol” no. 67.
- Kirson E.D., Dbaly V., Tovarys F., Vymazal J., Soustiel J.F., Itzakhi A. (2007), *Alternating Electric Fields Arrest Cell Proliferation in Animal Tumor Models and Human Brain Tumors*, „Proc Natl Acad Sci” no. 104.
- Kirson E.D., Gurvich Z., Schneiderman R., Dekel E., Itzakhi A., Wasserman Y. (2004), *Disruption of Cancer Cell Replication by Alternating Electric Fields*, „Cancer Res” no. 64.
- Kudowski J., Ludyn D., Przekwas M. (1997), *Energetyka a ochrona środowiska*, Warszawa.
- Li X., Zhang M., Bai L., Bai W. (2012), *Effects of 50 Hz Pulsed Electromagnetic Fields on the Growth and Cell Cycle Arrest of Mesenchymal Stem Cells: An in vitro Study*, „Electromagn Biol Med.” no. 31.
- Lin H., Blank M., Rossol-Haseroth K., Goodman R. (2001), *Regulating Genes with Electromagnetic Response Elements*, „J Cell Biochem.” no. 81.
- London S.J., Thomas D.C., Bowman J.D., Sobel E., Cheng T.C., Peters J.M. (1991), *Exposure to Residential Electric and Magnetic Fields and Risk of Childhood Leukemia*, „Am J Epidemiol.” no.134(9).
- Ohkubo T., Yamazaki J. (2012), *T-type Voltage-Activated Calcium Channel Cav3.1, but not Cav3.2, is Involved in the Inhibition of Proliferation and Apoptosis in MCF-7 Human Breast Cancer Cells*, „Int J Oncol.” no. 41.
- Palti Y. (1966), *Stimulation of Internal Organs by Means of Externally Applied Electrodes*, „J Appl Physiol” no. 21.
- Pawlowski P., Szutowicz I., Marszałek P., Fikus M. (1993), *Bioelectrorheological Model of the cell. 5. Electrodestruction of the Cellular Membrane in Alternating Electrical Field*, „Biophys J” no. 65.
- Polk C. (1995), *Therapeutic Applications of Low-Frequency Sinusoidal and Pulsed Electric and Magnetic Fields* [w:] J.D. Bronzino (red.), *The Biomedical Engineering Handbook*, Boca Raton, FL.
- Preston-Martin S. (1996), *Breast Cancer and Magnetic Fields*, „Epidemiology” no. 7.
- Repacholi M.H., Greenebaum B. (1999), *Interaction of Static and Extremely Low Frequency Electric and Magnetic Fields with Living Systems: Health Effects and Research Needs*, „Bioelectromagnetics” no. 20.
- Ruiz-Go’mez M.J., Martí’nez-Morillo M. (2005), *Enhancement of the Cell-Killing Effect of Ultraviolet-C Radiation by Short-Term Exposure to a Pulsed Magnetic Field*, „Int J Radiat Biol” no. 81.

- Sadeghipour R., Ahmadian S., Bolouri B., Pazhang Y., Shafieezadeh M. (2012), *Effects of Extremely Low-Frequency Pulsed Electromagnetic Fields on Morphological and Biochemical Properties of Human Breast Carcinoma Cells (T47D)*, „Electromagn Biol Med.” no. 31.
- Saliev T., Tachibana K., Bulanin D., Mikhalovsky S., Whitby R.D. (2014), *Bio-Effects of Non-Ionizing Electromagnetic Fields in Context of Cancer Therapy*, „Frontiers in Bioscience.” no. 6.
- Savitz D.A., Wachtel H., Barnes F.A., John E.M., Tvrdek J.E. (1988), *Case-Control Study of Childhood Cancer and Exposure to 60 Hz Magnetic Fields*, „Am J Epidemiol.” no. 128.
- Siemiński M. (1994), *Fizyka zagrożeń środowiska*, Warszawa.
- Silva C.P., Oliveira C.R., Lima M.C.P. (1996), *Apoptosis as a Mechanism of Cell Death Induced by Different Chemotherapeutic Drugs in Human Leukemic T-lymphocytes*, „Biochem Pharmacol” no. 51.
- Simko M. (2007), *Cell Type Specific Redox Status is Responsible for Diverse Electromagnetic Field Effects*, „Cur Med Chem.” no. 14.
- Stevens R.G., Davis S. (1996), *The Melatonin Hypothesis: Electric Power and Breast Cancer*, „Environ Health Perspect.” no. 104(suppl 1).
- Stevens R.G. (1987), *Electric Power Use and Breast Cancer: A Hypothesis*, „Am J Epidemiol.” no. 125(4).
- Stratton D., Lange S., Inal J.M. (2013), *Pulsed Extremely Low-Frequency Magnetic Fields Stimulate Microvesicle Release from Human Monocytic Leukaemia Cells*, „Biochem Bioph Res Commun.” no. 430.
- Taylor J.T., Huang L., Pottle J.E., Liu K., Yang Y., Zeng X. et al. (2008b), *Selective Blockade of T-type Ca<sup>2+</sup> Channels Suppresses Human Breast Cancer Cell Proliferation*, „Cancer Lett.” no. 267.
- Taylor J.T., Zeng X.B., Pottle J.E., Lee K., Wang A.R., Yi S.G. et al. (2008a), *Calcium Signaling and T-type Calcium Channels in Cancer Cell Cycling*, „World J Gastroenterol.” no. 14.
- Tokalov S.V., Gutzeit H.O. (2004), *Weak Electromagnetic Fields (50 Hz) Elicit a Stress Response in Human Cells*, „Environ Res.” no. 94.
- Tomenius L. (1986), *50-Hz Electromagnetic Environment and the Incidence of Childhood Tumors in Stockholm County*, „Bioelectromagnetics.” no. 7.
- Tomitsch J., Dechant E. (2015), *Exposure to Electromagnetic Fields in Households-Trends from 2006 to 2012*, „Bioelectromagnetics.” no. 36.
- Vijayalaxmi, Prihoda T.J. (2009), *Genetic Damage in Mammalian Somatic Cells Exposed to Extremely Low Frequency Electro-Magnetic Fields: A Meta-Analysis of 97 Publications (1990–2007)*, „Int J Radiat Biol.” no. 85.
- Wertheimer N., Leeper E. (1979), *Electrical Wiring Configurations and Childhood Cancer*, „Am J Epidemiol.” no. 109.
- Wolf F.I., Torsello A., Tedesco B., Fasanella S., Boninsegna A., D’Ascenzo M. et al. (2005), *50 Hz Extremely Low Frequency Electromagnetic Fields Enhance Cell Proliferation and DNA Damage: Possible Involvement of a Redox Mechanisms*, „Biochim Biophys Acta.” no. 1743.
- Yamaguchi S., Ogiue-Ikeda M., Sekino M., Ueno S. (2006), *Effects of Pulsed Magnetic Stimulation on Tumor Development and Immune Functions in Mice*, „Bioelectromagnetics” no. 27.

- Yan J., Dong L., Zhang B., Qi N. (2010), *Effects of Extremely Low-Frequency Magnetic Field on Growth and Differentiation of Human Mesenchymal Stem Cells*, „Electromagn Biol Med.” no. 29.
- Zhang X., Zhang H., Zheng C., LI C., Zhang X., Xiong W. (2002), *Extremely Low Frequency (ELF) Pulsed-Gradient Magnetic Fields Inhibit Malignant Tumour Growth at Different Biological Levels*, „Cell Biol Int” no. 26.
- Zimmerman J.W., Jimenez H., Pennison M.J., Brezovich I., Morgan D., Mudry A., Costa F.P., Barbault A., Pasche B. (2013), *Targeted Treatment of Cancer with Radiofrequency Electromagnetic Fields Amplitude-Modulated at Tumor-Specific Frequencies*, „Chin J Cancer” no. 32.
- Zimmerman J.W., Pennison M.J., Brezovich I., Yi N., Yang C.T. et al. (2012), *Cancer Cell Proliferation is Inhibited by Specific Modulation Frequencies*, „Br J Cancer” no. 106.
- Zimmerman U., Vienken J., Pivat G. (1981), *Rotation of Cells in an Alternating Electric Field: The Occurrence of a Resonance Frequency*, „Z Naturforsch” C 36.