Animal models of hypertension - revisited

Polak A.^{A-F*}, Harasim-Symbor E.^{C-F}, Chabowski A.^{C,E,F}

Department of Physiology, Medical University of Bialystok, Poland

A- Conception and study design; B - Collection of data; C - Data analysis; D - Writing the paper; E- Review article; F - Approval of the final version of the article; G - Other (please specify)

ABSTRACT

Nowadays, civilization diseases, such as hypertension, are one of the biggest global health problems. In 2017 the threshold for hypertension diagnosis was set at 130/80 mmHg, which resulted in its increased prevalence, reaching nearly 50% of the human population. Therefore, strategies for hypertension prevention and treatment have been recently extensively developing. Nonetheless, growing body of factors which can affect blood pressure and induce hypertension is constantly prompting researchers to conduct experiments in

DOI: 10.5604/01.3001.0012.1116

this field. For this purpose, animal models seem to be appropriate and necessary. The present report reviews current findings related to hypertension types and causes. It also presents the main guidelines for high blood pressure prevention and describes different experimental models introduced to be carried out in such studies.

Keywords: Dahl salt-sensitive rat, Deoxycorticosterone acetate-salt rat, Hypertension, Spontaneously hypertensive rat, Two-kidney oneclip model

*Corresponding author:

Agnieszka Polak, MSc, Zakład Fizjologii, Uniwersytet Medyczny w Białymstoku ul. Mickiewicza 2C, 15-222 Białystok, Poland, Tel.: +48 85 748 56 24; Fax: +48 85 748 55 85 e-mail: agn.polak@wp.pl

Received: 09.03.2018 Accepted: 07.05.2018 Progress in Health Sciences Vol. 8(1) 2018 pp 167-175 © Medical University of Białystok, Poland

INTRODUCTION

Hypertension is one of the risk factors for the development of cardiovascular diseases, which are responsible for more than 50% of the deaths globally [1]. The year 2017 brought changes in the scope of recommended threshold for hypertension diagnosis (Table 1). According to the latest American College of Cardiology (ACC) and American Heart Association (AHA) guidelines, high blood pressure should be treated at 130/80 mmHg rather than 140/90 mmHg [2]. Thereby, hypertension prevalence reaches half of the human population worldwide [2]. Therefore, it is not surprising that many of the recent experimental approaches are concentrated around this issue.

Table 1. New classification of blood pressure categories and hypertension stages recommended by the American College of Cardiology (ACC) and American Heart Association (AHA). SBP – systolic blood pressure, DBP – diastolic blood pressure

	Blood Pressure Categories			
	Normal	Elevated	Hypertension	
			Stage I	Stage II
SBP	< 120	120-129	130-139	≥ 140
(mm Hg)				
DBP	< 80	< 80	80-89	≥ 90
(mm Hg)				

CAUSES OF HYPERTENSION

I. Genetic predisposition

It is well known that many genes or gene variants may affect blood pressure (BP) [3]. To date, there are only several described single-gene mutations (e.g. in glucocorticoid-remediable aldosteronism, Gordon's or Liddle syndromes), which contribute to the development of hypertension [4]. However, these disorders are rarely observed. Usually, hypertension results from complex, polygenetic mutations as well as multiple single-nucleotide polymorphisms (SNPs) [3].

Genetic predisposition to elevation of BP is especially pronounced in primary hypertension, in which chronically increased BP has no identifiable cause. On the basis of studies carried out on twins, it has been proven that the heredity of hypertension reaches approximately 30-60% [5]. Moreover, family history of the hypertensive individuals also confirms relevant role of the genetic background in the etiology of hypertension [6].

II. Secondary hypertension

In contrast to the genetic basis of increased BP, secondary hypertension is caused by identifiable factors, which can explain occurrence of the elevated BP. However, this hypertension type affects only 10% of the adult patients [7]. Nevertheless, correct diagnosis and treatment may effectively decrease BP and prevent further organ damage.

1.Lifestyle factors

Environmental factors as well as an unhealthy lifestyle are often indicated as the major causes of hypertension. Interestingly, appropriate and balanced diet might play a crucial role in maintaining physiological BP values. Experimental researches have shown that among all dietary components, sodium and potassium consumption has the most pronounced influence on BP value [8-10]. It was demonstrated that excessive sodium intake positively correlates with increased BP [8,9] and this relationship enhances with age [9,11]. On the other hand, it was shown that reduction of salt intake decreases BP in the normotensive as well as hypertensive persons [8]. Although, there were minor differences between races in the aforementioned BP lowering effect, since Asian and black populations were more sensitive to low sodium diet compared to Caucasians. In contrast, increased potassium intake is linked to reduce BP value [12]. Moreover, there is evidence that high potassium consumption neutralizes sodium's influence on BP [10], which may be important in the prevention of hypertension.

Currently, there is discussion upon advantages and disadvantages of alcohol drinking and its connection with increased BP. A growing body of evidence indicates that chronic and high (i.e. >30g of ethanol/day) alcohol consumption is associated with hypertension [13]. Whereas, moderate alcohol consumption reduces BP and exhibits beneficial effects on the cardiovascular system [14-16]. Recommended "safe" ethanol amount for healthy man and nonpregnant women was set up to 20g/day and 10g/day, respectively [17]. Therefore, alcohol should be drunk in small doses in order to sustain its positive properties.

It is widely accepted that sedentary lifestyle and lack of adequate levels of physical activity may promote hypertension. Importantly, numerous studies indicate on the existence of a direct link between physical activity and the reduction of BP, which is most prominent in Caucasians [18-20]. Accordingly, even moderate activity, such as 20-minutes walk a day, can successfully prevent the development of hypertension [20]. Moreover, hypotensive benefits have been already noted for three 30-60-minutes exercise sessions per week at the intensity of 50-87% of maximal oxygen consumption [21]. Additionally, exercises diminish the elevation of BP with age, what provides the evidence that regular physical activity may reduce the risk of developing hypertension in the aging population [22].

What is interesting, it was found that BMI above 25 kg/m², which occurs in overweight and obese humans, is also related with increased risk for high BP [23,24]. Thus, maintaining a normal body weight, except for balanced diet and workout, is the main guideline of lifestyle modifications for prevention and treatment of hypertension [2].

2. Hypertension secondary to diseases

obstructive То date. sleep apnea, renovascular diseases, and primary aldosteronism seem to be the most common disorders, in which hypertension may coexist [2]. Obstructive sleep apnea is a chronic malfunction caused by complete or partial obstructions of the upper airways during sleeping, which induces episodes of apnea or hypopnea and results in hypoxemia and sleep disruptions [25]. Moreover, there is a positive correlation between an increased risk of hypertension and obstructive sleep apnea [26]. It is suggested that more than 80% of the adult patients with resistant hypertension are also affected by this disorder [27]. The exact mechanism underlying obstructive sleep apnea-related hypertension is still unknown. Nonetheless, there is a conception that the increased activity of the sympathetic nervous system and the renin-angiotensin system (RAS) is responsible for alternations in structure and functioning of blood vessels as well as blood pressure elevation [28].

Renovascular hypertension is caused by narrowing or blockage of the arteries supplying the kidneys [29]. For this reason renal perfusion pressure decreases and the juxtaglomerular cells secrete renin. Consequently, the activated RAS elevates retention of sodium and water causing an increase in BP [29]. Renal artery stenosis results mainly from atherosclerotic disease (90%) [30], however, nonatherosclerotic disorders (such as fibromuscular dysplasia) can also induce development of renal hypertension [30,31].

Primary aldosteronism (also known as primary hyperaldosteronism or Conn's syndrome) is commonly caused by adrenal hyperplasia, adrenal carcinoma [32] and rarely may be inherited [33]. In primary aldosteronism production of aldosterone is too high in comparison with plasma sodium concentration and is not suppressed by sodium loading [34]. Moreover, this excessive aldosterone production is not sensitive to the major regulators of its secretion, such as renin-angiotensin system or plasma potassium concentration [34]. Since primary aldosteronism causes increased renal sodium reabsorption and concomitant potassium excretion. 9-37% of patients with this disorder may also develop hypokalemia apart from hypertension [35].

ANIMAL MODELS OF HYPERTEN-SION

The animal models provide possibility not only to investigate the mechanisms involved in the pathogenesis of certain diseases, but also to screen potential therapies. Among several experimental models, rats are the most commonly used animals [36]. Moreover, since the etiology of human hypertension is heterogeneous, several types of animal models for hypertension assessment are introduced.

I. Animal models of primary hypertension

Interestingly, spontaneous development of high BP without any pharmacological or surgical intervention may occur in individual animals. This observation has contributed to the creation of genetic animal model of hypertension and enabled investigation of the genetic background of the disease. In 1963, Okamoto and Aoki introduced the first experimental model of primary hypertension, known as spontaneously hypertensive rat (SHR) [37]. The above researchers obtained a strain of rats with spontaneous hypertension by inbreeding Wistar-Kyoto rats with "naturally" abnormal high BP [37]. In this model, BP starts rising around 5-6th week of age and systolic blood pressure (SBP) reaches level of 180-200 mmHg in the mature rats [37]. It is suggested that at least 3 loci (on chromosomes 1, 3 and 4) are responsible for early development of hypertension in SHR. While gene detected on chromosome 10 promotes maintenance of higher BP values during aging in these animals [38]. Similarly to hypertensive patients, the SHRs develop cardiac hypertrophy and failure together with renal dysfunction. In spite of depressed endothelial-dependent relaxation response, these

animals do not exert major vascular problems, such as stroke, atherosclerosis or vascular thrombosis [37]. Interestingly, further-developed substrain named stroke-prone SHR [SHR-SP], apart from higher BP [SBP of about ~240 mmHg], exhibits severe vascular damages and increased incidences of deaths from a stroke compared to SHRs [39]. However, noteworthy advantage of this model is similarity of stroke course in SHR-SP and humans. This in turn, enables application of these animals in stroke studies, which is a common complication of hypertension in humans. On the other hand, there are experimental models, which manifest their predisposition to high BP under certain conditions. For example, in Dahl salt-sensitive strain, increased dietary sodium intake leads to severe (average SBP is over 200 mmHg) and fatal hypertension [40]. It has been proven that in this strain the above mentioned phenomenon is regulated bv angiotensin-converting enzyme and atrial natriuretic peptide receptor genes [40]. Interestingly, even though BP values in Dahl saltsensitive rats are higher compared to SHRs, the stage of cardiac hypertrophy is comparable in these two hypertension models [41,42]. However, it should be underlined that cardiac failure occurs earlier in Dahl salt-sensitive rats than in SHRs (4-5 vs 18 months of age) [41,42]. Moreover, renal changes in this strain are more severe and appear quicker in contrast to SHRs [43]. Recent knowledge progress and development of genetic engineering enable introduction of animals with overexpression or deletion of genes, which are involved in the regulation of BP. One of the first transgenic rat model was TGR(mRen2)27, in which overexpression of the mouse Ren2 renin gene led to hypertension [44]. Nonetheless, commonly used in the studies "knockout" animals, such as mice lacking the genes, which code ACE [45], angiotensin II type 1a receptor [46], endothelial synthase [47] or natriuretic peptide [48] also enable determining the function of particular genes by evaluating effects of their absence.

II. Animal models of secondary hypertension

1.Environmental models of hypertension

Apart from studies upon genetic factors, the growing body of research is focused on the influence of environment on BP. Interestingly, it was demonstrated that low temperature (around 5° C) may cause nearly 40% elevation of BP in animals within 3 weeks [49]. The above mentioned report is consistent with findings in humans, since people who chronically work in cold areas also develop hypertension [50]. Moreover, it was noticed that BP values in humans are higher in winter than in summer [51]. Furthermore, there are models, in which hypertension is generated by provoking stress in animals (e.g. using flash lights, loud noises or shaking) [52,53]. Additionally, high fat, sugar or salt diet can increase BP in animals and may be implemented as a method of hypertension induction [54,55]. Importantly, it is suggested that increased activity of the RAS, as well as the sympathetic nervous system, plays a pivotal role in the pathogenesis of described above environmental models of hypertension [55-58].

2. Renal hypertension

It is well known that renovascular disorders can cause an elevation of BP. Similar conditions may be triggered in animals by applying surgical procedures. There are two main methods, which are widely used for this purpose.

The first one, created in 1934 by Goldblatt et al. [59], is performed by a constriction of one or both renal arteries using a small clamp. To date, there are following Goldblatt's technique variants: one-kidney one-clip (1K1C; one renal artery is constricted and concomitantly the contralateral kidney is removed), two-kidney one-clip (2K1C; one renal artery is constricted and the contralateral kidney is left intact) and two-kidney two-clip (2K2C; aorta or both renal arteries are constricted) [60]. In contrast to other Goldblatt's models, in 1K1C hypertension only initial increase of BP is due to RAS activation. As a result of disruption in the functioning of the remaining kidney, no compensatory excretion of sodium and water is observed and greater volume of fluid is retained. Taking into consideration the mechanism of BP elevation, the 1K1C model represents volumerather than RAS-dependent type of hypertension [61]. Increased BP in the 2K1C model results from chronic hyperactivity of RAS, which is caused by artery constriction. Since one kidney is intact, it compensates impaired function of the second kidney and help sustain fluid and electrolyte balance in this model. Therefore, 2K1C model of hypertension is sensitive to RAS inhibition, but not to diuretics action [43]. In turn, the mechanism involved in the induction of the 2K2C hypertension is similar to the 2K1C model. However, more severe renal damages are observed in 2K1C rats, including acute renal failure and a high incidence of spontaneous stroke [62].

The second method of renovascular hypertension generation is based on procedures, which provoke renal parenchyma damages. It can be performed for instance by a renal mass reduction, which mimics chronic renal disease [63]. On the other hand, the external compression of the kidney e.g. by wrapping the kidney in cellophane, is often applied to obtain perinephritic fibrosis, which is formed after kidney transplantation [64]. Furthermore, microembolization-induced ischemia may be used to generate nephrosclerosis [65]. As high salt diet remarkably accelerates hypertension progression in the above mentioned animal models, it may be used in combination with surgical procedures to save time and costs of experiments [66].

3. Endocrinal hypertension

Pharmacological approaches for the induction of hypertension are also extensively used, e.g. in models of endocrine-related hypertension, such as deoxycorticosterone acetate (DOCA)-salt rats. In this model of secondary hypertension synthetic mineralocorticoid derivate - DOCA, combined with sodium chloride in unilateral nephrectomised rats produce volume overload hypertension [67]. The most important advantage of DOCA-salt hypertension is markedly depressed RAS activity, which enables its use in studies as an angiotensin-independent model [68]. It was found that DOCA-salt hypertension causes an elevation in the sympathetic nervous system activity [69]. Moreover, there is enhanced vasopressin secretion, which additionally increases renal water retention and leads to vasoconstriction [70]. Furthermore, DOCA-salt hypertension immediately progresses to severe hypertension (SBP < 200 mmHg) and cardiac hypertrophy. Therefore, it is useful model experimental investigation of all its for complications, especially those in the cardiovascular system [67]. It should be mentioned that other mineralocorticoids, such as aldosterone or glucocorticoids e.g. cortisol, can also initiate this type of hypertension [71,72].



Figure 1. Animal models of hypertension in experimental research. 1K1C – one-kidney one-clip, 2K1C – two-kidney two-clip, AT1a receptor – angiotensin II type 1a receptor, ACE – angiotensin-converting enzyme, DOCA- deoxycorticosterone acetate, SHR – spontaneously hypertensive rat, SHR-SP – stroke-prone spontaneously hypertensive rat, TGR(mREN2)27 – rats overexpressing the mouse Ren2 gene.

CONCLUSIONS

According to new guidelines for hypertension detection, nearly half of the human population may suffer from this disorder. Since cardiovascular diseases are the major cause of death globally, it is not surprising that strategies for treatment and prevention of hypertension are currently one of the most common area of research.

The present report describes the most popular animal models of hypertension as well as provides the information concerning novelties in the field of high blood pressure. We believe that this review will help understanding how important issue is hypertension. Moreover, we present comprehensive scope of experimental animal models, which are used for hypertension evaluation (Figure 1).

Conflicts of interest: None declared

Funding: This work was supported by the Medical University of Bialystok (grant number: N/ST/MN/17/003/1118).

REFERENCES

- Chen R, Dharmarajan K, Kulkarni VT, Punnanithinont N, Gupta A, Bikdeli B, Mody PS, Ranasinghe I. Most important outcomes research papers on hypertension. Circ Cardiovasc Qual Outcomes. 2013 Jul;6(4):e26-35.
- 2. Whelton PK, Carey RM, Aronow WS, Casey DE, Collins KJ, Dennison Himmelfarb C, DePalma SM, Gidding S, Jamerson KA, Jones DW, MacLaugghlin EJ, Muntner P, Ovbiagele B, Smith SC Jr, Spencer CC, Stafford RS, Taler SJ, Thomas RJ, Williams KA Sr, Williamson JD. Wright JT Jr 2017 ACC/AHA/AAPA/ABC/ACPM/AGS/APhA/A SH/ASPC/NMA/PCNA Guideline for the Prevention, Detection, Evaluation. and Management of High Blood Pressure in Adults: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. J Am Coll Cardiol. 2017 Nov.
- Padmanabhan S, Caulfield M, Dominiczak AF. Genetic and molecular aspects of hypertension. Circ Res. 2015 Mar;116(6):937-59.
- Lifton RP, Gharavi AG, Geller DS. Molecular mechanisms of human hypertension. Cell. 2001 Feb;104(4):545-56.
- 5. Kupper N, Ge D, Treiber FA, Snieder H. Emergence of novel genetic effects on blood pressure and hemodynamics in adolescence: the

Georgia Cardiovascular Twin Study. Hypertension. 2006 May;47(5):948-54.

- Lauer RM, Burns TL, Clarke WR, Mahoney LT. Childhood predictors of future blood pressure. Hypertension. 1991 Sep;18(3 Suppl):174-81.
- Calhoun DA, Jones D, Textor S, Goff DC, Murphy TP, Toto RD, White A, Cushman WC, White W, Sica D, Ferdinand K, Giles TD, Falkner B, Carey RM.Resistant hypertension: diagnosis, evaluation, and treatment: a scientific statement from the American Heart Association Professional Education Committee of the Council for High Blood Pressure Research. Circulation. 2008 Jun;117(25):e510-26.
- Graudal NA, Hubeck-Graudal T, Jurgens G. Effects of low sodium diet versus high sodium diet on blood pressure, renin, aldosterone, catecholamines, cholesterol, and triglyceride. Cochrane Database Syst Rev. 2017 Apr;4:CD004022.
- 9. Takase H, Sugiura T, Kimura G, Ohte N, Dohi Y. Dietary Sodium Consumption Predicts Future Blood Pressure and Incident Hypertension in the Japanese Normotensive General Population. J Am Heart Assoc. 2015 Jul;4(8):e001959.
- 10. Rodrigues SL, Baldo MP, Machado RC, Forechi L, Molina MeC, Mill JG. High potassium intake blunts the effect of elevated sodium intake on blood pressure levels. J Am Soc Hypertens. 2014 Apr;8(4):232-8.
- 11. Klag MJ, He J, Coresh J, Whelton PK, Chen JY, Mo JP, QIan MC, Mo PS, He GQ. The contribution of urinary cations to the blood pressure differences associated with migration. Am J Epidemiol. 1995 Aug;142(3):295-303.
- 12. Zhang Z, Cogswell ME, Gillespie C, Fang J, Loustalot F, Dai S, Carriquiry AL, Kuklina EV, Hong Y, Meritt R, Yang Q. Association between usual sodium and potassium intake and blood pressure and hypertension among U.S. adults: NHANES 2005-2010. PLoS One. 2013;8(10):e75289.
- Husain K, Ansari RA, Ferder L. Alcoholinduced hypertension: Mechanism and prevention. World J Cardiol. 2014 May;6(5): 245-52.
- Worm N, Belz GG, Stein-Hammer C. Moderate wine consumption and prevention of coronary heart disease. Dtsch Med Wochenschr. 2013 Dec;138(51-52):2653-7.
- 15. Brügger-Andersen T, Pönitz V, Snapinn S, Dickstein K, group Os. Moderate alcohol consumption is associated with reduced longterm cardiovascular risk in patients following a complicated acute myocardial infarction. Int J Cardiol. 2009 Apr;133(2):229-32.

- 16. Rimm EB, Klatsky A, Grobbee D, Stampfer MJ. Review of moderate alcohol consumption and reduced risk of coronary heart disease: is the effect due to beer, wine, or spirits. BMJ. 1996 Mar;312(7033):731-6.
- 17. Di Minno MN, Franchini M, Russolillo A, Lupoli R, Iervolino S, Di Minno G. Alcohol dosing and the heart: updating clinical evidence. Semin Thromb Hemost. 2011 Nov;37(8):875-84.
- Lesniak KT, Dubbert PM. Exercise and hypertension. Curr Opin Cardiol. 2001 Nov;16(6):356-9.
- Leary AC, Donnan PT, MacDonald TM, Murphy MB. The influence of physical activity on the variability of ambulatory blood pressure. Am J Hypertens 2000 Oct;13(10):1067-73.
- 20. Hayashi T, Tsumura K, Suematsu C, Okada K, Fujii S, Endo G. Walking to work and the risk for hypertension in men: the Osaka Health Survey. Ann Intern Med. 1999 Jul;131(1):21-6.
- 21. Halbert JA, Silagy CA, Finucane P, Withers RT, Hamdorf PA, Andrews GR. The effectiveness of exercise training in lowering blood pressure: a meta-analysis of randomised controlled trials of 4 weeks or longer. J Hum Hypertens 1997 Oct;11(10):641-9.
- 22. Carnethon MR, Gidding SS, Nehgme R, Sidney S, Jacobs DR, Liu K. Cardiorespiratory fitness in young adulthood and the development of cardiovascular disease risk factors. JAMA. 2003 Dec;290(23):3092-100.
- 23. Antza C, Stabouli S, Natsis M, Doundoulakis I, Kotsis V. Obesity-induced hypertension: new insights. Curr Pharm Des. 2017 Jun.
- 24. Dua S, Bhuker M, Sharma P, Dhall M, Kapoor S. Body mass index relates to blood pressure among adults. N Am J Med Sci. 2014 Feb; 6(2):89-95.
- 25. Parati G, Lombardi C, Hedner J, Bonsignore MR, Grote L, Tkacova R, Levy P, Riha R, Narkiewicz K, Mancia G, Bassetti C, WT; McNicholas European Respiratory Society; EU COST ACTION B26 members. Position paper on the management of patients with obstructive sleep apnea and hypertension: joint recommendations by the European Society of Hypertension, by the European Respiratory Society and by the members of European COST (Cooperation in Scientific and Technological research) ACTION B26 on obstructive sleep apnea. J Hypertens. 2012 Apr;30(4):633-46.
- 26. Marin JM, Agusti A, Villar I, Forner M, Nieto D, Carrizo SJ, - Barbé F, Vicente E, Wei Y, Nieto FJ, Jelic S. Association between treated and untreated obstructive sleep apnea and risk of hypertension. JAMA. 2012 May;307 (20):2169-76.

- 27. Muxfeldt ES, Margallo VS, Guimarães GM, Salles GF. Prevalence and associated factors of obstructive sleep apnea in patients with resistant hypertension. Am J Hypertens. 2014 Aug;27(8):1069-78.
- 28. Dopp JM, Reichmuth KJ, Morgan BJ. Obstructive sleep apnea and hypertension: mechanisms, evaluation, and management. Curr Hypertens Rep. 2007 Dec;9(6):529-34.
- 29. Textor SC. Secondary hypertension: renovascular hypertension. J Am Soc Hypertens 2014 Dec;8(12):943-5.
- 30. Weber BR, Dieter RS. Renal artery stenosis: epidemiology and treatment. Int J Nephrol Renovasc Dis. 2014;7:169-81.
- 31. Olin JW, Froehlich J, Gu X, Bacharach JM, Eagle K, Gray BH, Jaff MR, Kim ES, Mace P, Matsumoto AH, McBane RD, Kline-Rogers E, White CJ, Gornik HL. The United States Registry for Fibromuscular Dysplasia: results in the first 447 patients. Circulation. 2012 Jun;125(25):3182-90.
- 32. Schirpenbach C, Reincke M. Primary aldosteronism: current knowledge and controversies in Conn's syndrome. Nat Clin Pract Endocrinol Metab. 2007 Mar;3(3):220-7.
- 33. Brown MJ. Platt versus Pickering: what molecular insight to primary hyperaldosteronism tells us about hypertension. JRSM Cardiovasc Dis. 2012 Sep;1(6), doi: 10.1258/cvd.2012.012020.
- 34. Romero DG, Yanes Cardozo LL. Clinical Practice Guideline for Management of Primary Aldosteronism: What is New in the 2016 Update? Int J Endocrinol Metab Disord. 2016;2(3), 10.16966/2380-548X.129.
- 35. Mulatero P, Stowasser M, Loh KC, Fardella CE, Gordon RD, Mosso L, -Gomez-Sanchez CE, Veglio F, Young WF. Increased diagnosis of primary aldosteronism, including surgically correctable forms, in centers from five continents. J Clin Endocrinol Metab. 2004 Mar;89(3):1045-50.
- Dornas WC, Silva ME. Animal models for the study of arterial hypertension. J Biosci. 2011 Sep;36(4):731-7.
- Okamoto K, Aoki K. Development of a strain of spontaneously hypertensive rats. Jpn Circ J. 1963 Mar;27:282-93.
- 38. Mashimo T, Nabika T, Matsumoto C, Tamada T, Ueno K, Sawamura M, Ikeda K, Kato N, Nara Y, Yamori Y. Aging and salt-loading modulate blood pressure QTLs in rats. Am J Hypertens. 1999 Nov;12(11 Pt 1):1098-104.
- 39. Yamori Y, Horie R, Handa H, Sato M, Fukase M. Pathogenetic similarity of strokes in strokeprone spontaneously hypertensive rats and humans. Stroke. 1976 Jan-Feb;7(1):46-53.

- 40. Deng Y, Rapp JP. Cosegregation of blood pressure with angiotensin converting enzyme and atrial natriuretic peptide receptor genes using Dahl salt-sensitive rats. Nat Genet. 1992 Jul;1(4):267-72.
- 41. Inoko M, Kihara Y, Sasayama S. Neurohumoral factors during transition from left ventricular hypertrophy to failure in Dahl salt-sensitive rats. Biochem Biophys Res Commun. 1995 Jan;206(3):814-20.
- 42. Conrad CH, Brooks WW, Robinson KG, Bing OH. Impaired myocardial function in spontaneously hypertensive rats with heart failure. Am J Physiol. 1991 Jan;260(1 Pt 2):H136-45.
- 43. Pinto YM, Paul M, Ganten D. Lessons from rat models of hypertension: from Goldblatt to genetic engineering. Cardiovasc Res. 1998 Jul;39(1):77-88.
- 44. Mullins JJ, Peters J, Ganten D. Fulminant hypertension in transgenic rats harbouring the mouse Ren-2 gene. Nature. 1990 Apr;344(6266):541-4.
- 45. Esther CR, Howard TE, Marino EM, Goddard JM, Capecchi MR, Bernstein KE. Mice lacking angiotensin-converting enzyme have low blood pressure, renal pathology, and reduced male fertility. Lab Invest. 1996 May;74(5):953-65.
- 46. Sugaya T, Nishimatsu S, Tanimoto K, Takimoto E, Yamagishi T, Imamura K, Goto S, Imaizumi K, Hisada Y, Otsuka A. Angiotensin II type 1a receptor-deficient mice with hypotension and hyperreninemia. J Biol Chem. 1995 Aug;270(32):18719-22.
- 47. Ruetten H, Dimmeler S, Gehring D, Ihling C, Zeiher AM. Concentric left ventricular remodeling in endothelial nitric oxide synthase knockout mice by chronic pressure overload. Cardiovasc Res 2005 Jun;66(3):444-53.
- 48. Armstrong DW, Tse MY, O'Tierney-Ginn PF, Wong PG, Ventura NM, Janzen-Pang JJ, Matangi MF, Johri AM, Croy BA, Adams MA, Pang SC. Gestational hypertension in atrial natriuretic peptide knockout mice and the developmental origins of salt-sensitivity and cardiac hypertrophy. Regul Pept 2013 Sep;186:108-15.
- 49. Sun Z, Cade R, Morales C. Role of central angiotensin II receptors in cold-induced hypertension. Am J Hypertens 2002 Jan;15(1 Pt 1):85-92.
- 50. Donaldson GC, Robinson D, Allaway SL. An analysis of arterial disease mortality and BUPA health screening data in men, in relation to outdoor temperature. Clin Sci (Lond). 1997 Mar;92(3):261-8.
- 51. Brennan PJ, Greenberg G, Miall WE, Thompson SG. Seasonal variation in arterial

blood pressure. Br Med J (Clin Res Ed). 1982 Oct;285(6346):919-23.

- 52. Zimmerman RS, Frohlich ED. Stress and hypertension. J Hypertens Suppl. 1990 Sep;8(4):S103-7.
- 53. Buñag RD, Takeda K, Riley E. Spontaneous remission of hypertension in awake rats chronically exposed to shaker stress. Hypertension. 1980 May-Jun;2(3):311-8.
- 54. Kaufman LN, Peterson MM, Smith SM. Hypertension and sympathetic hyperactivity induced in rats by high-fat or glucose diets. Am J Physiol. 1991 Jan;260(1 Pt 1):E95-100.
- 55. Lee LC, Sasaki S, Inoue A, Fukuyama M, Nakamura Y, Oguro M, -Kawasaki S, Hayashi J, Takeda K, Yoshimura M. Salt increases blood pressure with biphasic changes in hypothalamic responsiveness in rats. J Cardiovasc Pharmacol. 1988 Aug;12(2):179-85.
- 56. Papanek PE, Wood CE, Fregly MJ. Role of the sympathetic nervous system in cold-induced hypertension in rats. J Appl Physiol (1985). 1991 Jul;71(1):300-6.
- 57. Coste SC, Qi Y, Brooks VL, McCarron DA, Hatton DC. Captopril and stress-induced hypertension in the borderline hypertensive rat. J Hypertens. 1995 Dec;13(12 Pt 1):1391-98.
- 58. Higashiura K, Ura N, Takada T, Li Y, Torii T, Togashi N, -Takada M, Takizawa H, Shimamoto K. The effects of an angiotensinconverting enzyme inhibitor and an angiotensin II receptor antagonist on insulin resistance in fructose-fed rats. Am J Hypertens. 2000 Mar;13(3):290-7.
- 59. Goldblatt H, Lynch J, Hanzal RF, Summerville WW. Studies on experimental hypertension : I. The production of persistent elevation of systolic blood pressure by means of renal ischemia. J Exp Med. 1934 Feb;59(3):347-79.
- 60. Doggrell SA, Brown L. Rat models of hypertension, cardiac hypertrophy and failure. Cardiovasc Res 1998 Jul;39(1):89-105.
- Brunner HR, Kirshman JD, Sealey JE, Laragh JH. Hypertension of renal origin: evidence for two different mechanisms. Science 1971 Dec;174(4016):1344-6.
- 62. Zeng J, Zhang Y, Mo J, Su Z, Huang R. Twokidney, two clip renovascular hypertensive rats can be used as stroke-prone rats. Stroke. 1998 Aug;29(8):1708-13; discussion 13-4.
- 63. Koletsky S, Goodsitt AM. Natural history and pathogenesis of renal ablation hypertension. Arch Pathol. 1960 Jun;69:654-62.
- 64. Page IH. A method for producing persistent hypertension by cellophane. Science 1939 Mar;89(2308):273-4.
- 65. Moore S, Mersereau WA. Microembolic renal ischemia, hypertension, and nephrosclerosis. Arch Pathol. 1968 Jun;85(6):623-30.

- 66. Ylitalo P, Hepp R, Möhring J, Gross F. Effects of varying sodium intake on blood pressure and renin-angiotensin system in subtotally nephrectomized rats. J Lab Clin Med. 1976 Nov;88(5):807-16.
- 67. Iyer A, Chan V, Brown L. The DOCA-Salt Hypertensive Rat as a Model of Cardiovascular Oxidative and Inflammatory Stress. Curr Cardiol Rev. 2010 Nov;6(4):291-7.
- Hilditch A, Hunt AA, Gardner CJ, Twissell DJ, Polley J, Travers A, Drew GM, Middlemiss D, Ross BC, Robertson MJ. Cardiovascular effects of GR117289, a novel angiotensin AT1 receptor antagonist. Br J Pharmacol. 1994 Jan;111(1):137-44.
- 69. Takeda K, Nakamura Y, Okajima H, Hayashi J, Kawasaki S, Lee LC, Sasaki S, Nakagawa M. Attenuated cardiovascular and sympathetic

nerve responses to aortic nerve stimulation in DOCA-salt hypertensive rats. J Hypertens. 1988 Jul;6(7):559-63.

- Ouchi Y, Yazaki Y, Tsai RC, Ashida T. Pressor response to vasopressin and norepinephrine in DOC-salt hypertensive and prehypertensive rats. Tohoku J Exp Med. 1988 Feb;154(2):125-33.
- 71. Knowlton AI, Loeb EN, Stoerk HC, White JP, Heffernan JF. Induction of arterial hypertension in normal and adrenalectomized rats given cortisone acetate. J Exp Med. 1952 Sep;96(3):187-205.
- 72. Garwitz ET, Jones AW. Aldosterone infusion into the rat and dose-dependent changes in blood pressure and arterial ionic transport. Hypertension. 1982 May-Jun;4(3):374-81.